Does basal PR gene expression in Solanum species contribute to non-specific resistance to Phytophthora infestans?

VIVIANNE G. A. A. VLEESHOUWERS^{1,3*}[†], WILLEM VAN DOOIJEWEERT¹, FRANCINE GOVERS^{2,3}, SOPHIEN KAMOUN^{2,3}[‡] and LEONTINE T. COLON¹

¹Plant Research International, Wageningen University and Research Center, P.O. Box 16, NL-6700 AA, Wageningen, The Netherlands, ²Laboratory of Phytopathology, Wageningen University and Research Center, Binnenhaven 9, NL-6709 PD, Wageningen, The Netherlands and ³Graduate School Experimental Plant Sciences, Wageningen, The Netherlands

(Accepted for publication May 2000)

Systemic acquired resistance (SAR) occurs in many plant species, including potato. SAR can be induced by various signals, but also basal levels of SAR may vary between plants. In *Arabidopsis* mutants, basal SAR levels positively correlate with pathogen resistance. Here we test whether in 13 wild *Solanum* clones and five potato cultivars, basal expression levels of SAR marker genes correlate with resistance to *Phytophthora infestans*. Most of the examined *Solanum* plants displayed significant and variable levels of race/ isolate-non-specific, partial resistance to five *P. infestans* isolates of diverse origin. Constitutive mRNA levels of the pathogenesis-related genes *PR-1*, *PR-2* and *PR-5* in non-infected leaves varied between the *Solanum* clones. However, no correlation between basal *PR* mRNA levels and resistance was observed at the genus level. In contrast, significant correlation was found at the species level in *S. arnezii* × *hondelmannii*, *S. microdontum*, *S. sucrense* and *S. tuberosum*. In *S. tuberosum* cultivars, the levels of *PR* gene expression were the highest in resistant Robijn, intermediate in partially resistant Première, Estima and Ehud, and the lowest in susceptible Bintje. These results suggest that constitutive expression of *PR* genes may contribute to nonspecific resistance to *P. infestans* in *Solanum*. Therefore, *PR* mRNAs could serve as molecular markers in potato breeding programs.

Keywords: Late blight; non-specific resistance; partial resistance; pathogenesis-related proteins; glucanase; osmotin; systemic acquired resistance; *Phytophthora infestans; Solanum arnezii* \times *hondelmannii; Solanum berthaultii; Solanum circaeifolium; Solanum microdontum; Solanum nigrum; Solanum sucrense; Solanum tuberosum; Solanum vernei;* wild *Solanum* species; potato.

INTRODUCTION

Resistance of plants to pathogens can be attributed to the action of various resistance mechanisms, each functioning at a certain level and specificity. One such mechanism is systemic acquired resistance (SAR), which generally follows a localized unsuccessful pathogen attack, involves an enhanced state of resistance to a broad spectrum of pathogens and is associated with an increased expression of genes encoding pathogenesis-related (PR) proteins

* To whom all correspondence should be addressed. E-mail: Vivianne.Vleeshouwers@medew.fyto.wau.nl

[†] Present address: Laboratory of Phytopathology, Wageningen University, Binnenhaven 9, NL-6709 PD Wageningen, The Netherlands.

[‡] Present address: Department of Plant Pathology, The Ohio State University, Ohio Agricultural Research and Development Center, Wooster, OH 44691, U.S.A.

Abbreviations used in text: IE, infection efficiency; LGR, lesion growth rate; PR, pathogenesis-related; SAR, systemic acquired resistance.

[30]. In other cases, pathogen attack is hampered by another type of resistance, called induced systemic resistance (ISR), which is not associated with increased PR gene expression [28].

SAR has been described in several plant species, including potato, but has best been documented in Arabidopsis thaliana. Several Arabidopsis mutants altered in SAR have been identified. For example, mutants in the npr1 gene (non-expresser of PR genes) fail to respond to SAR-inducing treatments, and are susceptible to the bacterial pathogen Pseudomonas syringae in contrast to wild-type plants [7, 13, 31]. Overexpression of npr1 led to enhanced resistance to P. syringae and the oomycete Peronospora parasitica in a dosage-dependent fashion since levels of npr1 mRNA, NPR1 protein, PR-1 mRNA and resistance were positively correlated in the transformants [8]. The *cpr* (constitutive expressers of PR genes) [6] and dnd1 (defense with no cell death) mutants [47] exhibit constitutive SAR-related phenotypes including elevated mRNA levels for genes encoding PR proteins. The analyses of these *Arabidopsis* mutants indicate that subtle mutations may affect basal SAR levels, and that higher resistance levels can be reached by manipulating basal levels of SAR.

In solanaceous plants, SAR has also been reported. In potato, SAR could be induced by treatment with hyphal wall components, unsaturated fatty acids and jasmonic acid, resulting in enhanced resistance to the late blight oomycete pathogen *Phytophthora infestans* (Mont.) de Bary [14, 9, 10]. In engineered potato plants containing a transgene encoding a bacterio-opsin proton pump from *Halobacterium halobium*, the expression of several *PR* genes was increased and resistance to *P. infestans* was enhanced [2]. Naturally occurring constitutive SAR was found in hybrids of tobacco and in certain tomato lines in which enhanced resistance to various pathogens was associated with constitutive *PR* gene expression [3, 46, 19].

In addition to realization of full-fledged SAR, the effects of overexpression of single PR genes on resistance have been described. Overexpression of PR-1 in tobaccoincreased resistance to Phytophthora parasitica and Peronospora tabacina [4], and overexpression of *PR-5* (osmotin) slightly enhanced resistance to *P. infestans* in potato [21, 27, R. Li and A. Pereira, pers. comm.] and Solanum commersonii [49]. PR-1-like proteins are conserved within the plant, fungal, vertebrate and invertebrate kingdoms. In animals these proteins function as venoms, allergens, or are implicated otherwise in defense [41]. The purified PR-1 protein from tomato and tobacco inhibited germination of P. infestans zoospores in vitro and lesion growth in vivo [25]. PR-2 and PR-3 encode glucanases and chitinases respectively, and these enzymes may play a role in cell wall degradation. However, oomycetes lack chitin in their cell wall and are not expected to be affected by chitinases. Actin-binding studies suggested that a basic chitinase and an osmotin-like protein might be involved in cytoplasmic aggregation, an important event in potato's cellular defense to P. infestans [36]. In addition, PR-5 proteins play a role in osmotic stress, freezing tolerance, permeabilization of fungal and oomycetal plasma membranes and pathogen resistance [45, 21, 1, 49].

P. infestans is a major pathogen of potato and tomato. In recent years, the severity of this disease has increased dramatically, and a more profound insight in the mechanisms of resistance to *P. infestans* is needed to develop novel control strategies. In the *P. infestans*—potato interaction, the most commonly studied type of resistance is race-specific resistance, which is governed by single dominant resistant genes (R genes). Unfortunately, race-specific resistance is only effective against certain strains of the pathogen, and is easily overcome by rapid evolution of the pathogen resulting in a lack of durability in the field. In contrast, race-non-specific resistance is effective against all known strains or races of the pathogen. It is thought to be based on multiple genes, may be durable,

and is generally of a partial nature. Several wild Solanum species possess varying levels of partial resistance to P. infestans [11, 44], and in old potato cultivars, such as cv. Robijn, partial resistance appeared to be durable [12]. In a previous study, we cytologically analysed Solanum species inoculated with P. infestans, and found that defense responses were always associated with the hypersensitive response (HR), a programmed cell death defense response of plants. In partially resistant clones, hyphal escape occurred and growing lesions were established [18, 43]. The growth rate of these lesions varied between different Solanum clones, indicating that defense mechanisms other than the HR operate at different levels in the different clones.

Even though a causal link between the accumulation of PR proteins and SAR has not always been established, a correlation between the timing of PR gene expression and the onset and duration of SAR is evident from many studies [30]. Therefore, measuring expression levels of PR genes is an appropriate method for determining levels of SAR [30, 22]. In this study, we determined the variation in basal mRNA levels of SAR marker genes (PR-1, PR-2 and PR-5) in Solanum plants, and tested whether resistance to P. infestans in Solanum species is associated with high basal levels of SAR.

MATERIALS AND METHODS

Plant material

The plant material used in this study is listed in Table 2. The origin of the plant material and the *in vitro* propagation was described previously [44]. Plants were grown under controlled conditions in climate chambers with a 16 h/8 h day/night regime at 18/15 °C and HPIT (Philips) illumination. The condition of different batches of plants used for the various resistance tests and for the *PR* gene expression analyses was comparable [44].

Phytophthora infestans

Phytophthora infestans isolates of different origins were used (Table 1). The propagation of the isolates and the preparation of inoculum was performed following standard procedures [44].

Resistance assessment

The resistance levels of 18 Solanum clones to six P. infestans isolates were determined using a routine resistance assay [44]. Detached leaves were spot-inoculated (10 μ l) with a zoospore suspension of 50 000 spores ml⁻¹, and incubated at high humidity in the dark. On the fourth, fifth and sixth day after inoculation, the largest length and width

Isolate	Clonal lineage	Race	Origin	Host of origin	Year	Mating type	Provided by
IPO-0	US-1	0	Unknown	Unknown	1987	Al	Lo Turkensteen, Plant Research Int., Wageningen, The Netherlands
90128	Unknown	1.3.4.6.7.8.10.11	Geldrop, The Netherlands	Potato	1990	A2	Francine Govers, Wageninger University, Wageningen, The Netherlands
Mex580	Unknown	Complex	Toluca Valley, Mexico	S. demissum	Unknown	Al	Bill Fry, Cornell University, Ithaca, U.S.A.
BIN-16	US-6	Complex	Northwestern U.S.A.	Unknown	1992	Al	Bill Fry, Cornell University, Ithaca, U.S.A.
ME93-2A	US-8	Complex	Maine, U.S.A.	Potato	1993	A2	Bill Fry, Cornell University, Ithaca, U.S.A.

TABLE 1. Phytophthora infestans isolates used in this study

(perpendicular on the length) of the lesions were measured. The ellipse area $(A = 1/4 \times \pi \times \text{length} \times \text{width})$ was calculated, and the lesions were divided into two groups, i.e. "no growing lesion" $(A \leq 16 \text{ mm}^2)$, or "growing lesion" $(A > 16 \text{ mm}^2)$. The infection efficiencies (IE) were calculated as the percentage of growing lesions. The areas of the "growing lesion" group were square root transformed, and the average lesion growth rate (LGR) was estimated by linear regression on time. The mean LGRs were analysed with REML using Genstat [16].

Southern and Northern blot analysis

DNA was isolated from leaves of *Solanum* plants and digested with *Eco*RV [*33*]. The DNA was electrophorized, transferred to Hybond-N⁺, and the Southern blot was hybridized with various probes. For expression analyses, leaf material (3rd, 4th and 5th fully developed leaf) from healthy, uninoculated plants was harvested and immediately frozen in liquid nitrogen. Two independent RNA isolation [*42*] series were carried out. For each RNA sample 15 μ g was loaded, electrophorized and transferred to Hybond-N⁺. The Northern blot was hybridized concurrently with the Southern blot with probes representing the *PR-1*, *PR-2*, *PR-5* and tubulin gene at 65, 60, 60 and 65 °C respectively, and the blots were washed at 1, 0.5, 0.5 and 1 × SSC stringency, respectively.

Messenger RNA levels were determined from the Northern blots using a Fujix Bio-Imaging Analyser (BAS 2000). The signals were quantified in photostimulated luminescence (PSL) per mm². To correct for slight differences in loading the signals for *PR* gene expression were normalized to the constitutively expressed tubulin signal.

DNA probes

PR gene members previously described to be correlated to resistance were selected. As DNA templates for probe synthesis the following fragments were used: a 400 bp EcoRI/KpnI fragment of *StPR1-1*, a *PR-1* cDNA clone from potato [39], a 1300 bp EcoRI/XhoI fragment of an acidic glucanase cDNA clone from tomato [40], a PCR fragment amplified on tobacco genomic DNA for *PR-5* [23], and a 1800 bp EcoRI/XhoI fragment from cDNA clone pFB19 encoding tubulin from potato.

RESULTS AND DISCUSSION

Specific and non-specific resistance in Solanum

To test the correlation between high levels of SAR and resistance to *P. infestans*, we first carefully determined the resistance levels of a set of 18 *Solanum* plants to five isolates from different clonal lineages, races, geographical origins, hosts of origin, years of isolation and mating types (Table 1). Various types of resistance were noted as illustrated by the mean LGRs and IEs data (Table 2).

Statistical analyses revealed a highly significant interaction (P < 0.001) between Solanum clones and P. infestans isolates, indicating that race-specific resistance occurs. In potato cv. Ehud (RI), race-specific resistance was evident since inoculation with race 0 strain IPO-0 resulted in complete resistance (IE = 0, LGR = 0), whereas inoculation with isolate 90128 (virulent on RI plants), resulted in high IEs and LGRs (Table 2). In Estima and Première (R10), the race-specific response was less pronounced, as a considerable percentage of growing lesions was noted in the interaction with IPO-0. This is in line with previous findings that R1 functions as a "strong" R gene and R10as a "weak" R gene [38, 43]. Statistical analyses of LGR

V. G. A. A. Vleeshouwers et al.

TABLE 2. Resistance levels of Solanum clones to P. infestans isolates IPO-0, 90128, Mex580, BIN-16 and ME93-2A

		IPO-0		90128		Mex580		BIN-16		ME93-2A	
Solanum	Clone		IE	LGR	IE	LGR	IE	LGR	IE	LGR	IE
Solanum berthaultii	ber-9	0.0	0	0.0	0	0.0	0	0.0	0	0.0	0
S. berthaultii	ber-11	1.6	25	2.4	10	1.9	18	0.0	0	0.0	0
S. arnezii × hondelmannii	axh-63	$2 \cdot 0$	60	1.9	95	$2 \cdot 2$	88	1.6	30	1.8	25
S. arnezii × hondelmannii	axh-72	2.9	88	3.3	75	2.6	65	2.3	10	1.8	5
S. circaeifolium ssp. circaeifolium	circl	0.0	0	0.0	0	0.7	3	0.0	0	0.6	25
S. microdontum	mcd-167	0.0	0	$3 \cdot 2$	45	1.8	5	1.5	3	0.0	0
S. microdontum	mcd-178	1.3	18	2.7	25	1.1	8	2.4	15	1.4	15
S. microdontum var. gigantophyllum	mcd-265	4.6	100	5.4	90	4.1	95	$3 \cdot 2$	100	$2 \cdot 0$	65
S. sucrense	scr-23	3.4	88	5.0	45	3.0	85	4.1	15	3.1	15
S. sucrense	scr-71	3.5	43	4.1	100	$2 \cdot 2$	55	$2 \cdot 1$	30	2.9	20
S. vernei	vrn-530	1.4	28	3.8	80	2.9	53	4.5	18	1.0	10
ABPT (30×33) hybrid	ABPT-44	2.3	13	2.6	80	$2 \cdot 0$	10	1.7	23	3.4	25
S. nigrum	ngr-SN18	0.0	0	0.0	0	0.0	0	0.0	0	0.0	0
S. tuberosum	Bintje	4.1	100	5.0	75	3.7	98	2.7	85	4.0	35
S. tuberosum	Ehud (RI)	0.0	0	$4 \cdot 2$	100	3.7	100	3.0	83	3.3	35
S. tuberosum	Estima $(R10)$	3.6	80	4.8	100	3.5	33	2.8	53	0.8	10
S. tuberosum	Première $(R10)$	$3 \cdot 1$	45	3.5	90	3.8	33	3.3	28	$4 \cdot 0$	15
S. tuberosum	Robijn	0.9	18	2.9	100	1.7	50	1.6	25	2.0	40

The resistances are expressed as mean lesion growth rate (LGR, in mm day⁻¹) and infection efficiency (IE, in percentage), and are based on two experiments for IPO-0, Mex580, BIN-16, and one experiment for 90128 and ME93-2A.

 $LSD_{LGR} = 1.3.$

 $LSD_{IE} = 24.$

and IE between Solanum clones and P. infestans isolates also revealed a highly significant interaction (P < 0.001)when cultivars bearing R1 and R10 were excluded, suggesting that novel undefined R genes may occur in the examined set of Solanum plants. However, strong isolate-specific resistance [43] comparable to R1 was not evident. In contrast, weaker isolate-specific responses were common. For example, S. microdontum-167 was more susceptible to isolate 90128 than to the other isolates. S. sucrense-23 was more often infected by IPO-0 and Mex580 (IE = 88 and 85 % respectively) than by other isolates (IE = 15-45%), although high LGRs were noted in all interactions. Potato cv. Estima and Première exhibited similar IEs and LGRs to isolates IPO-0, 90128, Mex580 and BIN-16, but Estima was remarkably resistant to isolate ME93-2A.

Low LGR values are considered as indicators of nonspecific, partial resistance [12]. In addition to isolatespecific responses, *Solanum* clones differed in LGRs independently of the isolate tested. For example, potato cv. Robijn showed lower LGR values than Bintje with all tested isolates. Cultivar Estima and Première, and to a lesser extent cv. Ehud, showed intermediate LGRs, indicating a certain level of partial resistance for these cultivars. In *S. microdontum*, clone 167 and 178 displayed lower LGRs than clone 265, and in *S. arnezii* × hondelmannii, clone 63 generally displayed slightly lower LGRs than clone 72, independently of the isolate. This suggests that most of the examined *Solanum* plants display significant and variable levels of race/ isolate-non-specific, partial resistance to five *P. infestans* isolates of diverse origin.

Occurrence of PR genes in Solanum

To check whether sufficient cross-hybridization occurs between the PR gene probes and the selected plants, a Southern blot containing genomic DNA from the different Solanum species was hybridized with probes from a potato PR-1 gene, a tomato PR-2 gene, and a tobacco PR-5 gene. As shown in Fig. 1, the three probes cross-hybridized with DNA from all Solanum clones. In all cases multiple hybridizing bands were detected revealing the presence of multi-gene families for PR-1, PR-2 and PR-5 in Solanum species. In addition, there was variation in signal intensity among the hybridizing bands, suggesting sequence diversity or differences in copy number among the family members. Multi-gene families have been reported for PR-2 in S. tuberosum [5], for PR-5in S. commersonii [48], and for PR-1, PR-2 and PR-5 in tobacco [**41**].

For each of the three PR genes, the hybridization pattern within the species was reasonably conserved. Between species however, hybridization patterns were quite diverse. For PR-2, a certain specificity can be noted, as the hybridization patterns of the closely related species

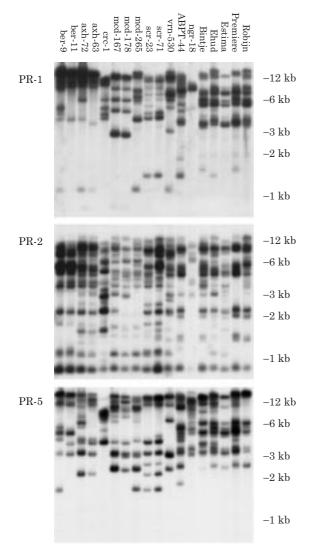


FIG. 1. Occurrence of *PR* genes in *Solanum* clones (for abbreviation of clones see Table 2). A Southern blot containing *Eco*RV digested genomic DNA isolated from *Solanum* clones was sequentially hybridized with a *PR-1* probe from potato, a *PR-2* probe from tomato, and a *PR-5* probe from tobacco.

S. berthaultii and S. arnezii × hondelmannii [35] were quite similar. In addition, the hybridization of the tomato PR-2probe to S. nigrum DNA was exceptionally weak, whereas hybridization of the same blot with the PR-1 and PR-5probes resulted in stronger signal intensities. This suggests that the PR-2 genes from S. nigrum are quite divergent from those of the other tested Solanum species. This is not surprising since S. nigrum is the most distantly related species in the examined set. S. nigrum is classified in the subgenus Solanum, whereas the other tested Solanum species and tomato belong to the subgenus Potatoe [34]. In summary, the Southern blot hybridizations showed that the heterologous PR probes are suitable for analysing expression of PR-1, PR-2 and PR-5 genes in the Solanum plants.

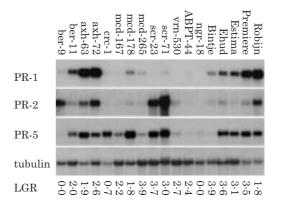


FIG. 2. Expression analyses of *PR* genes in uninoculated *Solanum* plants (for abbreviation of clones see Table 2). A Northern blot containing total RNA from *Solanum* clones was sequentially hybridized with *PR-1*, *PR-2*, *PR-5* and tubulin probes. The average LGRs (mm day⁻¹) (based on *P. infestans* isolates IPO-0, 90128, Mex580, BIN-16, ME93-2A) are indicated for each clone (LGR = 0 excluded).

Expression analysis of Solanum PR genes

To monitor constitutive SAR in the Solanum plants, the basal expression levels of the PR-1, PR-2 and PR-5 genes were determined in two independent experiments using uninoculated plants cultivated under defined conditions in growth chambers. The results of the first experiment are shown in Fig. 2. The autoradiographs showed that PR-1, PR-2 and PR-5 mRNAs were present at detectable levels in the majority of the tested plants. Interestingly, there is variation in PR mRNA levels between the different clones. In addition, within the entire Solanum set, the patterns of mRNA levels of the three PR genes were not identical. This is in line with the separation of *PR-1*, PR-2 and PR-5 gene activation pathways previously shown for Arabidopsis enhanced disease susceptibility mutants eds5 [29]. The absence of coordinated regulation between the different PR genes was also observed in salicylic acid induction deficient mutants sid1 and sid2, in which the pathway leading to PR-1 expression was blocked, whereas PR-2 and PR-5 were expressed at wild-type levels [24]. Thus unequal mRNA accumulation of the different PR genes could reflect different pathways of regulations for individual members of the three PRgene families, perhaps reflecting the wide range of physiological responses in which these proteins are involved [37, 26, 48].

PR mRNA levels were quite similar in the two experiments. However, in potato cv. Ehud and *S. vernei*-530, the mRNA levels observed for all three *PR* genes were higher in the second experiment compared to the first experiment. We assume that despite the precautions we took the Ehud and *S. vernei* plants in the second experiment may have been in a stressed state. In other experiments we noted high *PR-1* mRNA levels in plants grown in the greenhouse (data not shown), where they were exposed to heat and drought stress.

Does PR gene expression correlate with resistance?

To test whether isolate-non-specific, partial late blight resistance in Solanum is associated with a constitutive SAR, the obtained LGRs were compared to basal mRNA levels of PR genes. PR-1, PR-2 and PR-5 mRNA levels were quantified and normalized using tubulin mRNA levels as a reference. At the genus level, there was no indication for a correlation between non-specific resistance and PR expression levels. This may be explained by the different genetic background of the different plant species and the noted complexity of the three PR gene families. In contrast, significant correlation was observed at the species level. In S. arnezii \times hondelmannii, S. microdontum, S. sucrense and S. tuberosum, partially resistant clones exhibited higher levels of PR mRNA than more susceptible ones. In S. sucrense and S. arnezii \times hondelmannii, this correlation was evident for all three PR genes in the two independent experiments. In S. circaeifolium, S. vernei, ABPT and S. nigrum, only one clone was used, and thus correlations at the species level could not be tested. No correlation was observed between PR mRNA levels and resistance in S. berthaultii. However, the full resistance observed in clone 9 is HR-mediated [43] and suspected to operate through a novel R gene as recently observed for another accession of S. berthaultii [15].

Within S. tuberosum, the relationship between nonspecific resistance and PR mRNA levels was examined for the five tested cultivars (Fig. 3). Highly resistant cv. Robijn accumulated the highest levels of PR-1, PR-2 and PR-5 mRNA, whereas partially resistant Ehud, Estima and Première displayed intermediate levels, and susceptible Bintje very low levels. These results suggest a correlation between PR gene expression levels and resistance levels in potato. These results are consistent with those obtained with S. arnezii × hondelmannii, S. microdontum, and S. sucrense suggesting that an enhanced constitutive expression of SAR may be a component of the partial/non-specific resistance noted in Solanum species.

Concluding remarks

Basal SAR may function as an independent resistance mechanism, but more likely contributes to the complex network of defense reactions that take place following pathogen attack [17, 28]. Resistance in the Solanum-P. infestans interaction often exhibits a quantitative nature, which can be explained by the extent of an ambiguous HR. Per infection event, a fine balance between invading hyphae and plant cells exhibiting an HR, determines whether infection will be aborted [43]. Basal level of SAR may increase the sensitivity of plant

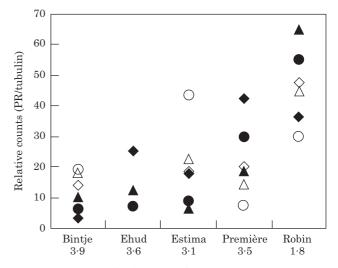


FIG. 3. Messenger RNA levels of *PR* genes and isolate-nonspecific resistance in *S. tuberosum. PR-1* (\bullet) *PR-2* (\blacktriangle) and *PR-5* (\bullet) mRNA levels from uninoculated leaves from potato cultivars Bintje, Ehud, Estima, Première and Robijn were quantified on a phospho-imager, and the PR signals were normalized using the tubulin mRNA level as reference. Messenger RNA levels were determined in two independent experiments (first experiment closed, second open symbols); the data from Ehud in the second experiment were excluded. On the X-axis, average LGRs (mm day⁻¹, see Table 2) determined after inoculation with *P. infestans* isolates IPO-0, 90128, Mex580, BIN-16, ME93-2A are indicated for each cultivar (LGR = 0 excluded).

cells to HR elicitation [32], or may slow pathogen invasion by creating physiological conditions that limit pathogen growth.

The identification of molecular markers linked to partial resistance to P. infestans in Solanum species, is of great value for potato late blight resistance breeding. High levels of PR mRNAs could serve as useful molecular markers for screening breeding populations and germplasm. Assays based on quantitative reverse transcriptase (RT-PCR) of PR genes could be developed to assist potato breeders and geneticists in identifying promising genotypes and could supplement other assays based on quantitative trait loci (QTL) [20]. Using marker genes for known resistance mechanisms such as SAR may provide a novel prospective for marker-assisted breeding.

We would like to thank Jan van Kan (Laboratory of Phytopathology, Wageningen University) for providing the *PR-2* probe, Andy Pereira (Plant Research International, Wageningen) for providing the *PR-5* probe, Paul Keizer (Plant Research International, Wageningen) for help with the statistical analyses, and Pierre de Wit (Laboratory of Phytopathology, Wageningen University) for critically reading of the manuscript. This work was financed by Stichting Stimulering Aardappelonderzoek, Zeist, The Netherlands.

REFERENCES

- Abad LR, Durzo MP, Liu D, Narasimhan ML, Reuveni M, Zhu JK, Niu XM, Singh NK, Hasegawa PM, Bressan RA. 1996. Antifungal activity of tobacco osmotin has specificity and involves plasma membrane permeabilization. *Plant Science* 118: 11–23.
- Abad MS, Hakimi SM, Kaniewski WK, Rommens CMT, Shulaev V, Lam E, Shah DM. 1997. Characterization of acquired resistance in lesion mimic transgenic potato expressing bacterio-opsin. *Molecular Plant-Microbe Interactions* 10: 635–645.
- Ahl Goy P, Felix G, Metraux JP, Meins F Jr. 1992. Resistance to disease in the hybrid Nicotiana glutinosa × Nicotiana debneyi is associated with high constitutive levels of beta-1,3-glucanase, chitinase, peroxidase and polyphenoloxidase. Physiological and Molecular Plant Pathology 41: 11-21.
- 4. Alexander D, Goodman RM, Gut Rella M, Glascock C, Weymann K, Friedrich L, Maddox D, Ahl Goy P, Luntz T, Ward E, Ryals J. 1993. Increased tolerance to two oomycete pathogens in transgenic tobacco expressing pathogenesis-related protein 1a. Proceedings of the National Academy of Sciences of the United States of America 90: 7327-7331.
- Beerhues L, Kombrink E. 1994. Primary structure and expression of mRNAs encoding basic chitinase and 1,3beta-glucanase in potato. *Plant Molecular Biology* 24: 353–367.
- Bowling SA, Guo AL, Cao H, Gordon AS, Klessig DF, Dong XN. 1994. A mutation in *Arabidopsis* that leads to constitutive expression of systematic acquired resistance. *Plant Cell* 6: 1845–1857.
- Cao H, Bowling SA, Gordon AS, Dong XN. 1994. Characterization of an *Arabidopsis* mutant that is nonresponsive to inducers of systemic acquired resistance. *Plant Cell* 6: 1583–1592.
- Cao H, Li X, Dong XN. 1998. Generation of broadspectrum disease resistance by overexpression of an essential regulatory gene in systemic acquired resistance. *Proceedings of the National Academy of Sciences of the United States of America* 95: 6531–6536.
- Cohen Y, Gisi U, Mosinger E. 1991. Systemic resistance of potato plants against *Phytophthora infestans* induced by unsaturated fatty acids. *Physiological and Molecular Plant Pathology* 38: 255–264.
- Cohen Y, Gisi U, Niderman T. 1993. Local and systemic protection against *Phytophthora infestans* induced in potato and tomato plants by jasmonic acid and jasmonic methyl ester. *Phytopathology* 83: 1054–1062.
- Colon LT, Budding DJ. 1988. Resistance to late blight (*Phytophthora infestans*) in ten wild *Solanum* species. *Euphytica* Suppl.: 77–86.
- Colon LT, Turkensteen LJ, Prummel W, Budding DJ, Hoogendoorn J. 1995. Durable resistance to late blight (*Phytophthora infestans*) in old potato cultivars. *European Journal of Plant Pathology* 101: 387–397.
- Delaney TP, Friedrich L, Ryals JA. 1995. Arabidopsis signal transduction mutant defective in chemically and biologically induced disease resistance. Proceedings of the National

Academy of Sciences of the United States of America **92**: 6602–6606.

- Doke N, Ramirez AV, Tomiyama K. 1987. Systemic induction of resistance in potato plants against *Phytophthora infestans* by local treatments with hyphal wall components of the fungus. *Journal of Phytopathology* 119: 232–239.
- Ewing EE, Simko I, Smart CD, Bonierbale MW, Mizubuti ESG, May GD, Fry WE. 2000. Genetic mapping from field tests of qualitative and quantitative resistance to *Phytophthora infestans* in a population derived from *Solanum tuberosum* and *Solanum berthaultii*. *Molecular Breeding* 6: 25-36.
- 16. **Genstat 5 Committee.** 1987. *Genstat 5 Reference Manual*. Oxford: Clarendon Press.
- Graham TL, Graham MY. 1999. Role of hypersensitive cell death in conditioning elicitation competency and defense potentiation. *Physiological and Molecular Plant Pathology* 55: 13–20.
- Kamoun S, Huitema E, Vleeshouwers VGAA. 1999. Resistance to oomycetes: a general role for the hypersensitive response? *Trends in Plant Science* 4: 196–200.
- Lawrence CB, Joosten MHAJ, Tuzun S. 1996. Differential induction of pathogenesis-related proteins in tomato by *Alternaria solani* and the association of a basic chitinase isozyme with resistance. *Physiological and Molecular Plant Pathology* 48: 361–377.
- Leonards-Schippers C, Gieffers W, Schafer Pregl R, Ritter E, Knapp SJ, Salamini F, Gebhardt C. 1994. Quantitative resistance to *Phytophthora infestans* in potato: a case study for QTL mapping in an allogamous plant species. *Genetics* 137: 67–77.
- 21. Liu D, Raghothama KG, Hasegawa PM, Bressan RA. 1994. Osmotin overexpression in potato delays development of disease symptoms. *Proceedings of the National Academy of Sciences of the United States of America* 91: 1888-1892.
- Maleck K, Dietrich RA. 1999. Defense on multiple fronts: How do plants cope with diverse enemies? *Trends in Plant Science* 4: 215–219.
- Melchers LS, Sela-Buurlage MB, Vloemans SA, Woloshuk CP, van Roekel JSC, Pen J, van den Elzen PJM, Cornelissen BJC. 1993. Extracellular targeting of the vacuolar tobacco proteins AP24, chitinase and beta-1,3-glucanase in transgenic plants. *Plant Molecular Biology* 21: 583–593.
- Nawrath C, Metraux JP 1999. Salicylic acid inductiondeficient mutants of *Arabidopsis* express PR-2 and PR-5 and accumulate high levels of camalexin after pathogen inoculation. *Plant Cell* 11: 1393–1404.
- 25. Niderman T, Genetet I, Bruyere T, Gees R, Stintzi A, Legrand M, Fritig B, Mosinger E. 1995. Pathogenesisrelated PR-1 proteins are antifungal: isolation and characterization of three 14-kilodalton proteins of tomato and of a basic PR-1 of tobacco with inhibitory activity against *Phytophthora infestans*. *Plant Physiology* **108**: 17–27.
- Ori N, Sessa G, Lotan T, Himmelhoch S, Fluhr R. 1990. A major stylar matrix polypeptide (sp41) is a member of the pathogenesis-related proteins superclass. *The EMBO Journal* 9: 3429–3436.
- Pereira A, Sandbrink H, Wolters P, van Enckevort E, van der Vossen E, Vleeshouwers VGAA, Colon LT, Stiekema W. 1998. Identification and engineering of disease resistance genes in potato. Abstract Book 5th International Symposium on the Molecular Biology of the Potato. Bogensee: Germany.

- Pieterse CMJ, van Loon LC. 1999. Salicylic acidindependent plant defence pathways. *Trends in Plant Science* 4: 52–58.
- Reuber TL, Plotnikova JM, Dewdney J, Rogers EE, Wood W, Ausubel FM. 1998. Correlation of defense gene induction defects with powdery mildew susceptibility in Arabidopsis enhanced disease susceptibility mutants. *Plant Journal* 16: 473–485.
- Ryals JA, Neuenschwander UH, Willits MG, Molina A, Steiner HY, Hunt MD. 1996. Systemic acquired resistance. *Plant Cell* 8: 1809–1819.
- 31. Shah J, Tsui F, Klessig DF. 1997. Characterization of a salicylic acid insensitive mutant (sai1) of Arabidopsis thaliana, identified in a selective screen utilizing the SA inducible expression of the tms2 gene. Molecular Plant-Microbe Interactions 10: 69–78.
- 32. Shirasu K, Nakajima H, Rajasekhar VK, Dixon RA, Lamb C. 1997. Salicylic acid potentiates an agonist dependent gain control that amplifies pathogen signals in the activation of defense mechanisms. *Plant Cell* 9: 261–270.
- Shure M, Wessler S, Fedoroff N. 1983. Molecular identification and isolation of the *Waxy* locus in maize. *Cell* 35: 225-233.
- 34. Spooner DM, Anderson GJ, Jansen RK. 1993. Chloroplast DNA evidence for the interrelationships of tomatoes, potatoes, and pepinos (Solanaceae). *American Journal of Botany* 80: 676–688.
- Spooner DM, Castillo TR. 1997. Reexamination of series relationships of South American wild potatoes (Solanaceae: Solanum sect. Petota): evidence from chloroplast DNA restriction site variation. *American Journal of Botany* 84: 671–685.
- Takemoto D, Furuse K, Doke N, Kawakita K. 1997. Identification of chitinase and osmotin-like protein as actin-binding proteins in suspension-cultured potato cells. *Plant and Cell Physiology* 38: 441–448.
- 37. Tornero P, Gadea J, Conejero V, Vera P. 1997. Two PR-1 genes from tomato are differentially regulated and reveal a novel mode of expression for a pathogenesis related gene during the hypersensitive response and development. Molecular Plant-Microbe Interactions 10: 624–634.
- Turkensteen LJ. 1987. Interactions of R genes in breeding for resistance of potatoes against *Phytophthora infestans*. In *Planning conference on fungal diseases of the potato*. Lima, Peru: CIP, 57–73.

- 39. van 't Klooster JW, Vleeshouwers VGAA, Kamoun S, Govers F. 1999. Characterization of a cDNA encoding a pathogenesis-related protein PR-1 from potato (Solanum tuberosum) (Accession No. AJ250136). Plant Physiology 121: 1384.
- 40. van Kan JAL, Joosten MHAJ, Wagemakers CAM, van den Berg-Velthuis GCM, de Wit PJGM. 1992. Differential accumulation of mRNAs encoding extracellular and intracellular PR proteins in tomato by virulent and avirulent races of *Cladosporium fulvum*. *Plant Molecular Biology* 20: 513-527.
- 41. van Loon LC, van Strien EA. 1999. The families of pathogenesis-related proteins, their activities, and comparative analysis of PR-1 type proteins. *Physiological and Molecular Plant Pathology* 55: 85–97.
- 42. Verwoerd TC, Dekker BMM, Hoekema A. 1989. A small scale-procedure for the rapid isolation of plant RNAs. *Nucleic Acids Research* 17: 2362.
- 43. Vleeshouwers VGAA, van Dooijeweert W, Govers F, Kamoun S, Colon LT. 2000. The hypersensitive response is associated with host and nonhost resistance to *Phytophthora infestans. Planta* **210**: 853–864.
- 44. Vleeshouwers VGAA, van Dooijeweert W, Keizer LCP, Sijpkes L, Govers F, Colon LT. 1999. A laboratory assay for *Phytophthora infestans* resistance in various *Solanum* species reflects the field situation. *European Journal of Plant Pathology* 105: 241–250.
- 45. Woloshuk CP, Meulenhoff JS, Sela Buurlage M, van den Elzen PJM, Cornelissen BJC. 1991. Pathogeninduced proteins with inhibitory activity toward *Phytophthora infestans. Plant Cell* **3**: 619–628.
- 46. Yalpani N, Shulaev V, Raskin I. 1993. Endogenous salicylic acid levels correlate with accumulation of pathogenesis-related proteins and virus resistance in tobacco. *Phytopathology* 83: 702–708.
- 47. Yu IC, Parker J, Bent AF. 1998. Gene-for-gene disease resistance without the hypersensitive response in Arabidopsis dnd1 mutant. Proceedings of the National Academy of Sciences of the United States of America 95: 7819–7824.
- Zhu B, Chen THH, Li PH, Zhu BL. 1995. Activation of two osmotin-like protein genes by abiotic stimuli and fungal pathogen in transgenic potato plants. *Plant Physiology* 108: 929–937.
- 49. Zhu B, Chen THH, Li PH, Zhu BL. 1996. Analysis of lateblight disease resistance and freezing tolerance in transgenic potato plants expressing sense and antisense genes for an osmotin-like protein. *Planta* 198: 70–77.

42